

Introduction

Rapid diagnostics is required in cases with respiratory failure for clinical decision making regarding isolation and antiviral therapy.^{1,2} Techniques like immune-chromatographic test (ICT) and direct immunofluorescence assay (DFA) have lower sensitivities and specificities than molecular diagnostic assays, but have the advantage of quick turnaround times and ease-of-use. Here, we evaluated the performance of an automated, easy to use, sample-to-answer system, which performs an Influenza A/B virus (fluA/fluB), respiratory syncytial virus (RSV) and sample processing control (SPC) multiplex RT-PCR of 1-12 samples within 2 hours.

Methods

The performance of the fluA/fluB/RSV assay on the ARIES (Luminex), a system using MultiCode technology (a probe-free real-time RT-PCR method with melting curve confirmation), was evaluated using published laboratory developed automated real-time RT-PCR assays (LDA) for fluA, fluB, RSV-A and RSV-B^{3,4}.

Analytical performance of the FluA/FluB/RSV assay (ARIES, Luminex):

- Genotype inclusivity**: 16 avian (H1-H16) and 33 human fluA strains, 3 fluB strains and the two RSV (A/B) strains.
- Analytical specificity**: 40 high positive non-fluA/fluB/RSV-viruses
- Analytical sensitivity**: 0.5 log dilution series of A/H1N1p2009 B/Yamagata, RSV-A and RSV-B compared to LDA assays.
- Linearity**: 0.5 log dilution series of A/H1N1p2009 B/Yamagata, RSV-A and RSV-B compared to LDA assays.
- Repeatability**: 35 replicates of a control positive for fluA, fluB and RSV in different runs.

Clinical performance: compared to both LDA ± ICT (BinaxNOW influenza A/B and RSV test) ± DFA using selected (pretreated), -80°C stored, respiratory tract samples from 2006 until 2015 (retrospective) and prospective testing of original respiratory tract samples from December 2015 onwards.

Results

Genotype inclusivity

*All fluA, fluB and RSV-A/B strains tested for analytical performance evaluation were detected. External lysis with MPLC lysisbuffer (Roche) of avian and highly pathogenic fluA strains yielded correct results.

*No aspecific reactions with non fluA/fluB/RSV high positive controls were identified.

Analytical sensitivity

ARIES fluA/fluB/RSV assay was less sensitive for fluA (0.5 log), RSV-A (1 log), RSV-B (2 log) and for FluB (2.5 log) compared to LDA

Repeatability

Replicates of a positive process control (PPC, n=35, figure 1)

	fluA		RSV		SPC	fluB	
	Ct	Tm °C	Ct	Tm °C	Tm °C	Ct	Tm °C
Average	34.1	84.0	33.6	75.8	78.3	36.6	79.8
STDEV	0.9	1.7	1.1	0.1	0.3	1.1	0.1
%CV	2.6	2.0	3.2	0.2	0.4	3.0	0.1

Figure 1: Results repeatability

Concentration of fluB in PPC, was close to the ARIES limit of detection, and tested positive 32 out of 35 (91.4%). T- and F-tests gave no significant difference (>0.05).

Linearity

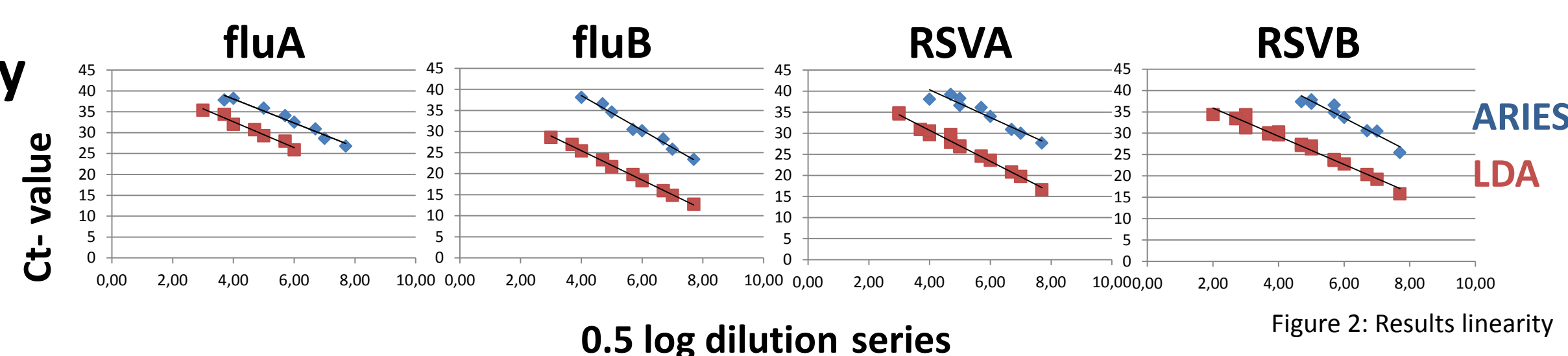


Figure 2: Results linearity

Robustness

1.8% of the cassettes failed during operation (pre-RUO and RUO).

1.1% of the cassettes failed during operation (RUO only).

Clinical performance

In total, 227 samples were included in the clinical performance evaluation. Sample type distribution (figure 3):

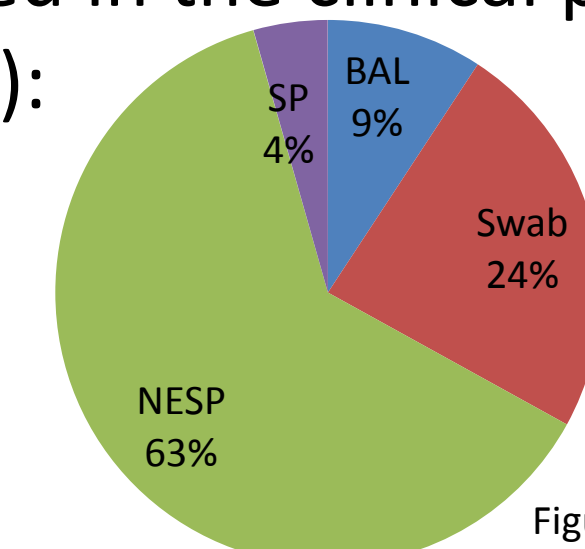


Figure 3: sample type distribution

17.9% tested positive for fluA, 14.2% for fluB and 42.7% for RSV, (RSV-A, 24.5% and RSV-B 20.4%) in both LDA and ARIES.

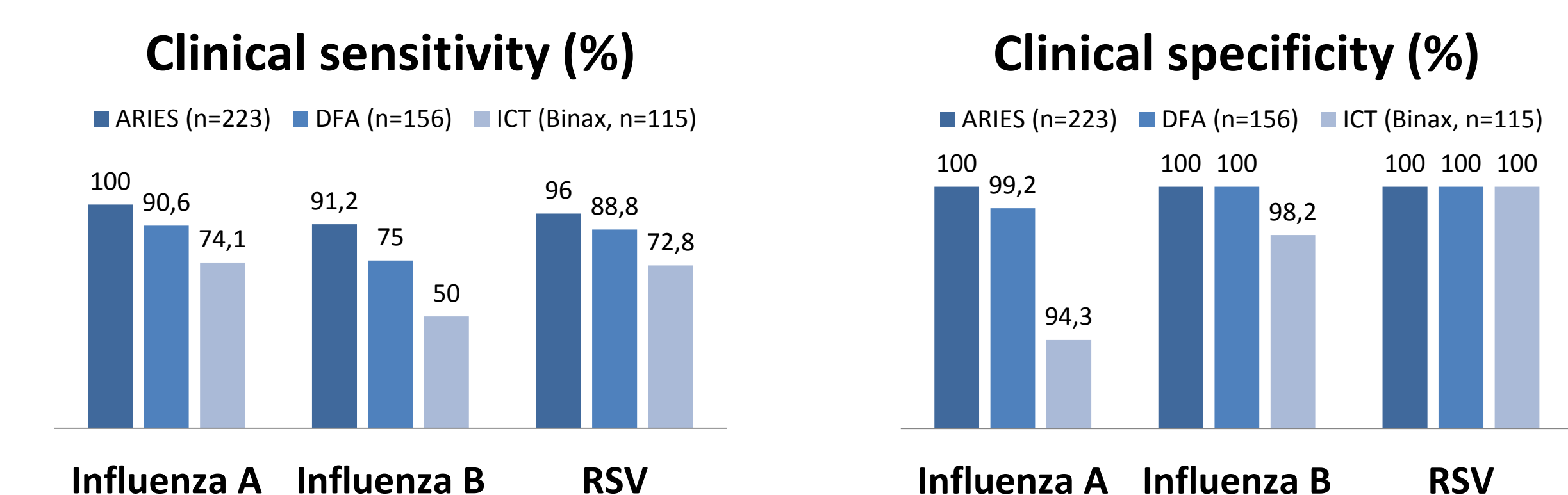


Figure 4: Clinical sensitivity and specificity

*Confirmed discrepant results were found in 7 samples (3 fluB and 4 RSV-A), which tested positive in LDA and negative in ARIES (3.1%, LDA Ct values 27.9 - 33.9).

*If compared to the DFA (n=156) and ICT (n=116), ARIES detected 17 (10.6%; 3 fluA, 5 fluB, 9 RSV) and 32 (28.1%; 7 fluA, 3 fluB, 22 RSV) more samples respectively, all confirmed by LDA (Ct range 16.5-31.6).

*ARIES has a higher clinical sensitivity and specificity compared to DFA and ICT (figure 4)

Conclusion

1. The ARIES influenza A/B/RSV assay is a specific and rapid molecular assay
2. Although analytically the ARIES is less sensitive for fluB and RSV-A and RSV-B than the LDA assays, the performance in clinical samples is comparable to LDA and better than those of the established rapid assays.

References:

- 1 Blaschke AJ et al, *Journal of the pediatric infectious diseases society*, Vol3, No.2, (2014)112-118
- 2 Dunn et al. *Diagnostic microbiology and infectious disease* 79 (2014) 10-13
- 3 Hoek RA et al. *Scand J Infect Dis.* 45 (2013) 65-69
- 4 Dewhurst-Maridor G et al. *Journal of Virological Methods* 120 (2004) 41-49

COI: The cassettes for this study were provided by Luminex Corp.

This study was approved by the local Medical ethical committee under MEC-2015-475

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